fact that an endocyclic double bond in a mediumsized ring will partially relieve the strain due to crowding of hydrogen atoms.

During the equilibration of methylenecyclodecane, trans-1-methylcyclodecene was formed initially at a rate only slightly slower than the *cis* isomer. This result indicates that the presumed carbonium ion precursor can lose a proton to give *cis* and trans endocyclic olefins with approximately equal ease. After the concentration of the trans isomer had reached a maximum value of 37% of the total olefin mixture, it decreased slowly to the equilibrium value of 0.5%. ARTHUR C. COPE

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RECEIVED APRIL 27, 1959

SOLVATION EFFECTS AND THE BAKER-NATHAN SEQUENCE

Sir:

In view of current interest in reëxamining the origin of the Baker–Nathan effect,¹ we are prompted to report some kinetic and thermodynamic measurements we have made at 25.06° with *p*-methylbenzyl chloride and *p*-*t*-butylbenzyl chloride, as solutions in methanol and as pure liquids.

First-order specific rate constants $(k, \text{ in sec.}^{-1})$ for the production of acid from solutions of the benzyl chlorides have been established as 3.06 ± 0.01 $\times 10^{-7}$ and $2.76 \pm 0.01 \times 10^{-7}$ for the *p*-methyl and *p*-*t*-butyl compounds, respectively. Thus, in this system, the Baker–Nathan sequence of substituent effects is followed with $k_{p\text{-methyl}} > k_{p\text{-}t\text{-butyl}}$.

By application of the dynamic vapor pressure technique,² and ultraviolet spectroscopic analysis, partial pressures (p_0 , in mm.) of the pure benzyl chlorides have been measured, the values being 0.18 \pm 0.01 and 0.017 \pm 0.001 for the *p*-methyl and *p*-*t*butyl compounds, respectively. Application of the same techniques to solutions of the benzyl chlorides has yielded Henry's law constants (*H*, in mm. mole/kg. solvent) of 0.19 \pm 0.01 and 0.026 \pm 0.002 for the *p*-methyl and *p*-*t*-butyl compounds, respectively, invariant with concentration over the range 0.100-0.00100 molal.

If one defines solvation energy as the free energy of the transformation, pure compound \rightarrow hypothetical 1 molal solution (as 2.303 $RT \log H/\dot{p}_0$),³ the solvation energies (in cal./mole) of the benzyl chlorides are $+30 \pm 30$ and $+250 \pm 50$ for the *p*methyl and *p*-*t*-butyl compounds, respectively.

 Conference on Hyperconjugation, Indiana University, June 2-4, 1958; Tetrahedron, 5, 105 (1959).

(2) A. L. Bacarella, A. Finch and D. Grunwald, J. Phys. Chem., 60, 573 (1956).

(3) This definition of solvation energy, although arbitrary, is analogous to that for the Gibbs free energy of mixing (E. A. Guggenheim, "Thermodynamics." Interscience Publishers, Inc., New York, N. Y., 1957, p. 242) which is a measure of the total change in free energy associated with a solution phenomenon. Like the latter, it has the advantage of yielding values for solvation effects which are insensitive to absolute volatilities, and hence significant for comparisons involving different compounds. Thus, for all substances which form ideal solutions, regardless of the vapor pressures of the pure substances, this definition yields identical solvation energies of 2.303 $RT \log X$, where N is the mole fraction of the solute in the hypothetical 1 molal solution.

Alternatively, solvation effects may be expressed in terms of the ratio of the experimental vapor pressure to that calculated from Raoult's law; values less than unity indicate enhanced interactions between solute and solvent over those in the pure components, and values greater than unity indicate reduced interactions.⁴ This ratio $(H/0.0310 \ p_c)$, for the hypothetical 1 molal solutions of benzyl chlorides, is 34 ± 2 and 49 ± 4 for the *p*-methyl and *p*-*t*-butyl compounds, respectively. By either criterion, it is apparent that, in this system, *p*-*t*-butyl-benzyl chloride is less strongly solvated than *p*-methylbenzyl chloride.

To our knowledge, these are the first measurements of solvation effects in a system which undergoes chemical reaction according to the Baker-Nathan sequence. The results support the contention⁵ that the Baker-Nathan sequence is associated with differential solvation effects rather than C-H hyperconjugation.

We wish to thank the National Science Foundation for its support of this work under grant number G-5116.

(4) W. J. Moore, "Physical Chemistry," Prentice-Hall, Inc., Englewood Cliffs, N. J., 1955, p. 135.

(5) W. M. Schubert, J. M. Craven, R. G. Minton and R. B. Murphy, *Tetrahedron*, **5**, 194 (1950), and earlier papers by W. M. Schubert. GEORGE HERBERT JONES LABORATORY

DEPARTMENT OF CHEMISTRY UNIVERSITY OF CHICAGO CHICAGO 37, ILLINOIS ROBERT A. CLEMENT JAMSHID N. NAGHIZADEH

RECEIVED APRIL 22, 1959

A NEW HYPOTENSIVE STEROID ALKALOID FROM CONOPHARYNGIA PACHYSIPHON

Sir:

In the course of our plant screening program we had occasion to prepare an extract of the roots of *Conopharyngia pachysiphon* (Apocynaceae) obtained from Trinidad.¹ This extract was found by our Macrobiology Division to exert considerable hypotensive activity when tested intravenously in dogs. However, this action was accompanied by a marked respiratory depressant effect. Chromatography of the crude extract on a silicic acid column effected a complete separation of these actions and permitted the crystallization of the pure hypotensive factor, m.p. 259–260°. Microanalysis indicated the formula C27,H45NO6 HCl. Treatment of the hydrochloride with ammonia yielded the free base, m.p. $285-288^{\circ}$, infrared absorption bands (Nu-jol) at 3528 cm. $^{-1}$ (OH); 3404, 3270 cm. $^{-1}$ (bonded OH, NH); 1595 cm. $^{-1}$ (NH of NH₂); multiple strong bands in 1000–1100 cm.⁻¹ region (C–O– \hat{C}). Acetylation with acetic anhydride in pyridine formed a pentaacetate, $C_{37}H_{55}NO_{11}$, m.p. $202-203^{\circ}$, infrared bands at 1750 cm.⁻¹ (O-acetyl); 1650 cm.⁻¹ (N-acetyl). Mild alkaline hydrolysis of the pentaacetate yielded a product, m.p. 269–271°, C29H47NO7, which still showed an infrared band indicative of N-acetyl, whereas the 1750 cm.⁻¹ band had disappeared completely. Therefore, it seemed evident that the pentaacetate contained four Oacetyl and one N-acetyl, a fact confirmed by acetyl determination. That a primary amine was origi-

(1) This material was very kindly collected and identified by Prof. F. J. Simmonds, Imperial College of Tropical Agriculture, St. Angustine, Trinidad, B.W.L, to whom we are most grateful. nally present was further demonstrated by a Van Slyke amino nitrogen assay.

Acid hydrolysis yielded two aglycons which were separated by chromatography; I, m.p. 111–115°, $C_{21}H_{38}N$, ultraviolet absorption at 229–230 mµ (ϵ 15,260); 234 mµ (ϵ 16,700), 243 mµ shld. (ϵ 12,190); II, m.p. 168–173°, $C_{21}H_{35}NO$. Compound II has now been identified as 20 α -amino-3 β -hydroxy-5-pregnene² and, therefore, I corresponds to 20 α -amino-3,5-pregnadiene. The sugar fraction of the hydrolysis was shown to be D-glucose by formation of its osazone, by paper chromatography and by its oxidation to potassium gluconate and subsequent formation of the characteristic aldobenzimidazole.³ From the above evidence and from other considerations it seemed quite probable that the new steroid alkaloid was 20 α -amino-3 β -hydroxy-5-pregnene β -D-glucoside. This has been confirmed by synthesis.

 20α -Amino- 3β -hydroxy-5-pregnene was converted to 3β -hydroxy- 20α -trifluoroacetamido-5pregnene, m.p. 199–201°. Reaction with acetobromoglucose gave the 3β -D-glucoside acetate, m.p. $200-205^{\circ}$. Alkaline hydrolysis removed the blocking groups and the resulting base, m.p. $285-287^{\circ}$, yielded a hydrochloride, m.p. $257-259^{\circ}$, which was identical in every respect with isolated hypotensively active material. Several derivatives and compounds related to this substance have been investigated and these together with more complete details of the above experiments will be reported later.

(2) P. L. Julian, E. W. Meyer and H. C. Printy, THIS JOURNAL, 70, 887 (1948). See also V. Cerny, L. Lobler and F. Sorm, *Coll. Czech.*, 22, 76 (1957), for the stereochemistry of the C-20 amines.

(3) S. Moore and K. P. Link, J. Biol. Chem., 133, 293 (1940).

RESEARCH DEPARTMENT DANIEL DICKEL CIBA PHARMACEUTICAL PRODUCTS, INC. ROBERT LUCAS SUMMIT, N. J. H. B. MACPHILLAMY RECEIVED MAY 7, 1959

Sir:

DISSOCIATION OF γ -GLOBULIN

Reaction of human γ -globulin with sulfhydryl compounds, sulfite, or performic acid resulted in marked diminution in the sedimentation coefficient and molecular weight. Efficient reduction required the presence of denaturing agents. Because the products were insoluble after removal of the denaturing agent, molecular weights were determined in urea solutions¹ by the Archibald principle²

$$q_{\rm a} \equiv \frac{RT(\partial c/\partial r)_{\rm ra}}{\omega^2 r_{\rm a}} = M_{\rm app} \left(1 - \bar{\rm v}\rho\right) \left[(c_{\rm a} - c_{\rm 0}) + c_{\rm 0} \right]$$

R is the gas constant, *T* the absolute temperature, $(\partial c/\partial r)_{1_a}$ the concentration gradient at the meniscus r_a , ω the constant angular velocity, M_{app} the apparent molecular weight of solute calculated as if its activity coefficient were unity, \bar{v} the partial specific volume, ρ the density of the solvent, c_a the concentration at r_a , and c_0 the initial concentration. Plotting q_a against $(c_a - c_0)$ yields $M_{app}(1 - \bar{v}\rho)$ as the slope of a least squares line. This allows calculation of a molecular weight average which

(1) R. Trautman and C. F. Crampton, Abstracts of Papers, 130th Meeting, American Chemical Society, Sept. 1956, 9-C; THIS JOURNAL, in press. initially emphasizes the heavier components of a polydisperse solution.³

Fifteen milligrams of human γ -globulin was reduced with 5 ml. of 0.1 M β -mercaptoethylamine-HCl in 6 M urea at room temperature for four hours. Some samples then were dialyzed against a large volume of 6 M urea that was 0.02 M in iodoacetamide. The products appeared as a polydisperse peak in the ultracentrifuge. Corrected for the density and viscosity of urea, the s^{0}_{20w} of reduced iodoacetamide-treated γ -globulin was 2.3S. In Table I are the measured values of $M_{app}(1 - \bar{\nu}\rho)$ and M_{app} values calculated assuming $\bar{\nu} = 0.74.^4$ Simultaneous determinations of $M_{app}(1 - \bar{\nu}\rho)$ with solutions in D₂O and H₂O that were 6 M in urea and 0.1 M in β -mercaptoethylamine-HCl, allowed estimation of $\bar{\nu} = 0.71$. Using this value, and $\rho = 1.097$ for the H₂O solution yields $M_{app} = 42,000$.

Table ${f I}$

Effect of Various Reagents on $M_{app}(1 - \bar{v}\rho)$ Values of Human γ -Globulin

(Fraction II of Cohn, Lederle lot C 543a)

Solvent	$M_{app}(1 - \tilde{v}\rho) \pm \text{standard} $ deviation	$M_{ m app}$
0.2 M KCl	$(4.8 \pm 0.1) \times 10^4$	192,000
6 M urea + 0.2 M KCl	$(3.0 \pm 0.3) \times 10^4$	158,000
$0.1 \ M \ MEA^b + 0.2 \ M$	$(3.5 \pm 0.1) \times 10^4$	140,000
K.Cl		
$0.1 \ M \text{ MEA} + 6 \ M$	$(0.93 \pm 0.07) \times 10^4$	48,000
urea $+ 0.2 M$ KCl		
Reduced in 8 M urea +	$(0.92 \pm 0.05) \times 10^4$	48,000
$0.1 \ M$ MEA, next		
dialyzed against 6 M		
urea $+ 0.02 M$ iodo-		
acetamide then $6~M$		
urea + $0.2 M$ KCl		

^a This sample contained a small amount of heavy material sedimenting faster than the main 7S component. ^b MEA = β -mercaptoethylamine·HCl.

A pathological macroglobulin reduced in 6 Murea had a value of 41,000 for $M_{\rm app}$. Macroglobulins have been found⁵ to dissociate in mercaptoethanol solutions without urea to sub-units of about the same molecular weight as normal γ -globulin.

Reaction of human γ -globulin with sulfite⁶ in urea⁷ yielded a water soluble aggregated S-sulfoprotein. In tris-(hydroxymethyl)-aminomethane buffer, pH 8, made 6 M in urea, the smallest non-dialyzable component had a value of 0.81×10^4 for $M_{\rm app}$ $(1 - \bar{\nu}\rho)$. Performic acid oxidation⁸ of human γ -globulin gave a product with $M_{\rm app} = 32,000$ that was slightly soluble in citrate buffer, pH 10.9, in the absence of urea.

These findings suggest that human γ -globulin contains subunits linked at least in part by disulfide

(3) D. A. Yphantis, J. Phys. Chem., in press.

(4) J. L. Oncley, G. Scatchard and A. Brown, J. Phys. Colloid Chem., 51, 184 (1947).

(5) H. F. Deutsch and J. I. Morton, J. Biol. Chem., 231, 1107 (1958).

(6) J. M. Swan, Nature, 180, 643 (1957).

(7) J. F. Pechère, G. H. Dixon, R. H. Maybury and H. Neurath, J. Biol. Chem., 233, 1364 (1958).

(8) C. H. W. Hirs, *ibid.*, **219**, 611 (1956)

⁽²⁾ R. Trautman, Biochim. et Biophys. Acta, 28, 417 (1958).